

## Technical Bulletin #C0030: gCOSY with CapNMR

*This bulletin describes how to use the CapNMR probe for gCOSY ( $^1\text{H}$ - $^1\text{H}$ ) on a Varian console/system.*

Begin by loading an appropriate 'Proton' sequence. Make sure to use the appropriate power levels (see Figure 1). The solenoid microcoil in the CapNMR probe requires less transmitter power than traditional probes. The standard pulse width (pw90) is calibrated at installation for a transmitter power of 45 dB (tpwr=45).

Parameter	Varian Range	Bruker Range
Lock power	1 to 8	-55 to -45
Lock gain	34 to 48	110 to 130
Transmitter power level (dB)	40 to 45	15 to 20
90° Transmitter pulse width (µsec)	2.5 to 5	2.5 to 5
90° Decoupling pulse width (µsec)	5 to 12	7 to 12

Figure 1. Appropriate power levels for the CapNMR Probe

Once there is a good Proton spectrum, type 'gCOSY' into the command prompt and this will load the gCOSY pulse sequence. You can view the pulse sequence with a dps command. The sequence should look like the sequence in Figure 2.

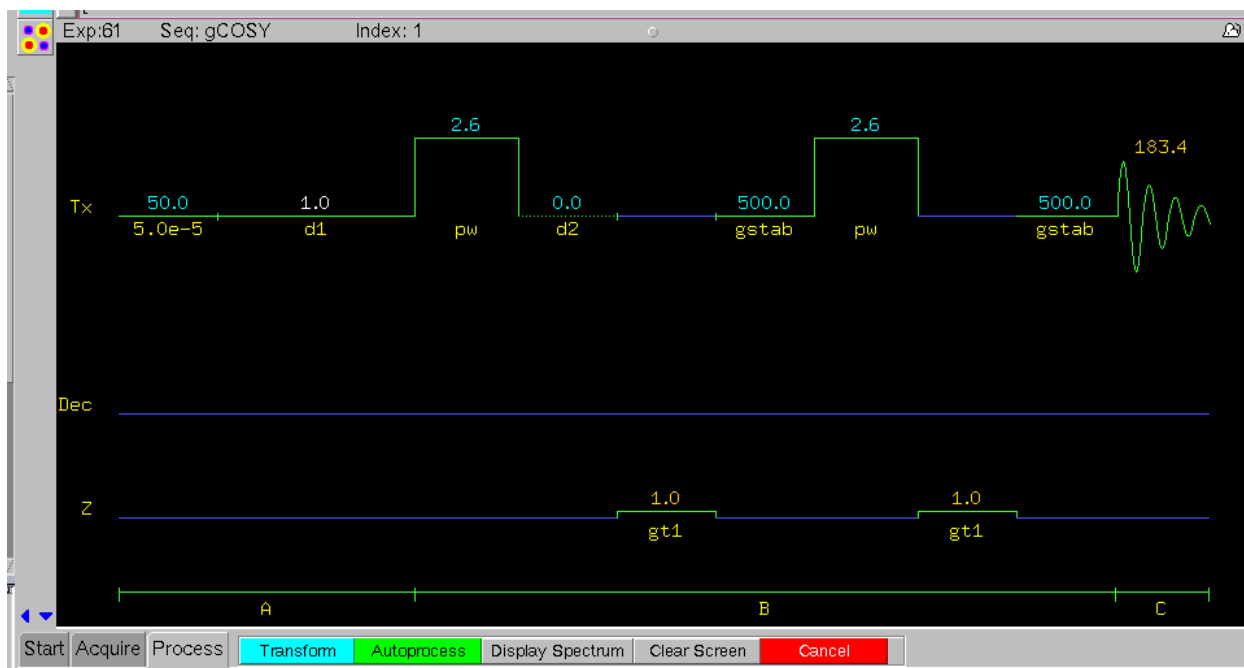


Figure 2. gCOSY Pulse Sequence

However, the pulse width is probe/console specific so your pulse width (pw) value should be different than the 2.6 value shown in Figure 2 and Figure 3. The pw90 for proton was calibrated with an array of the pw parameter in a Proton sequence and for tpwr=45, the pw90 for this probe was 2.6 microseconds.

Here is an example of the parameter set used for the gCOSY pulse sequence. In order to view this window, click on Process and select Text Output and then type 'dg'.

	ACQUISITION	TRANSMITTER	GRADIENTS	PROCESSING
seqfil	gCOSY	tn	H1	gzlvl1 888 sb -0.092
sw	5584.6	sfrq	599.763	gt1 0.001000 sbs not used
at	0.183	tof	-311.8	gstab 0.000500 fn 2048
np	2048	tpwr	45	hsglvl 888 2D PROCESSING
ss	16	pw	2.650	hsqt 0.005000 sb1 -0.023
dl	1.000	DECOUPLER		FLAGS sbs1 not used
nt	1	dn	C13	hs nn procl lp
ct	1	dm	nnn	sspul n fnl 2048
2D ACQUISITION		SPECIAL		SAMPLE
swl	5584.6	temp	not used	date Sep 14 2006
ni	128	spin	not used	solvent CDC13
PRESATURATION		gain	54	sample
satmode	nnn			

Figure 3. gCOSY Parameters for the CapNMR Probe

**IMPORTANT: GRADIENT POWER LEVELS**

The CapNMR probe has a much stronger z-gradient than most standard tube probes. Most common tube probes are 50-80 gauss/cm at 10 amps while the CapNMR is commonly 350-400 gauss/cm at 10 amps. The default gzlvl1 and hsglvl values from Varian are 2000. You may have seen these values as high as 4500 for some tube probes. However, due to a more efficient gradient coil, not as much gradient power (gzlvl) is required for the same gradient field.

**A standard gradient value (gzlvl1 & hsglvl) for the CapNMR probe is 700 to 900.**

These gradient and homospoil gradient values are automatically calculated from the gcal value in the probe file. The gcal value is calibrated at installation with a profile sequence, which performs a z gradient echo, and is saved in the probe file. Make sure your probe is set correctly (check with 'probe?') and that the 'probegcal' value is correct.

*Sample and Data Set:*

The sample used was 10% (v/v) 2-ethyl-1-indanone in CDCl<sub>3</sub>. Pure 2-ethyl-1-indanone is a clear, brown liquid with a T1 relaxation of approximately 0.8 seconds. This data set was taken on a **5-μL of 10% 2-ethyl-1-indanone (519 μg; 3.2 μmol; v/v in CDCl<sub>3</sub>)**, which was injected and then pushed into a 5 microliter flowcell with more CDCl<sub>3</sub> solvent. This data was taken on a Varian Inova at 600 MHz with a TXI (Triple Inverse <sup>1</sup>H, <sup>2</sup>H, <sup>13</sup>C, <sup>15</sup>N) CapNMR probe with a 5 microliter enhanced flowcell in 3 minutes.

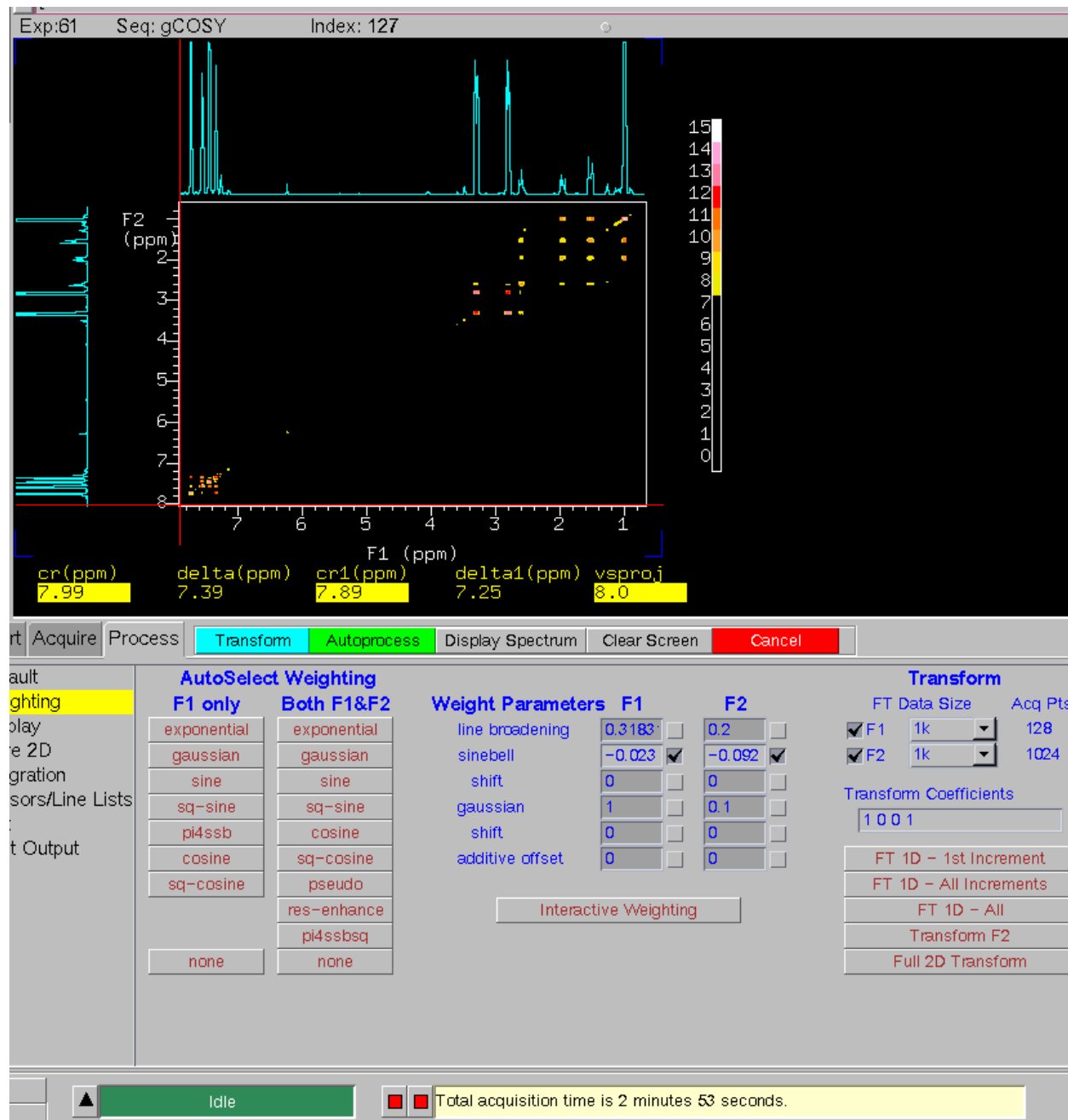


Figure 4. gCOSY Spectrum from 10% 2-ethyl-1-indanone

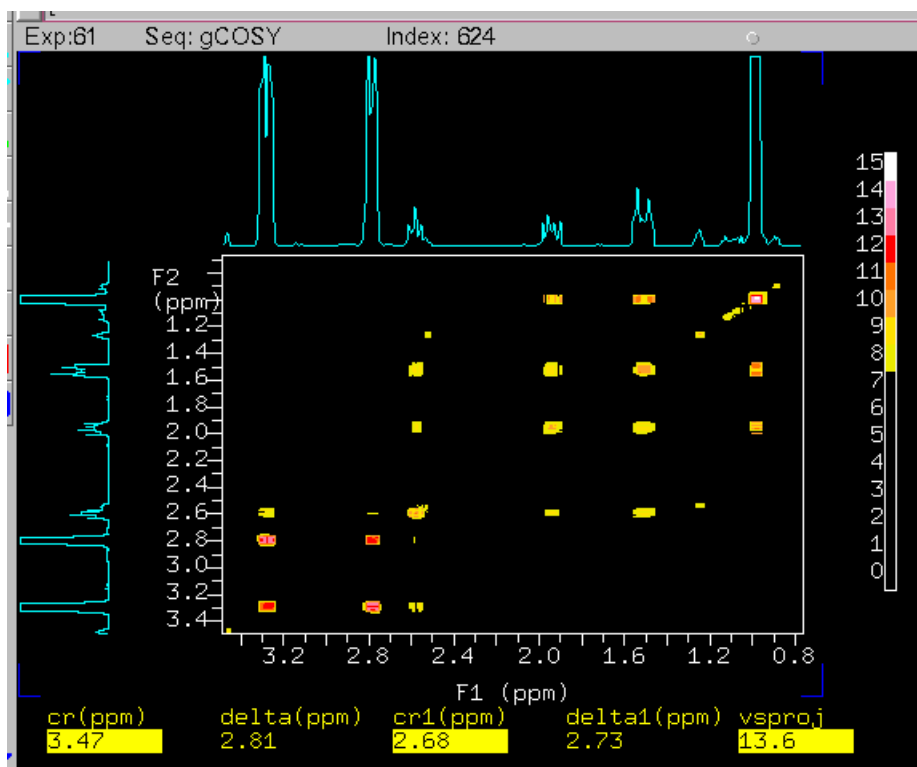


Figure 5. gCOSY Spectrum (Aliphatic Region) from 10% 2-ethyl-1-indanone

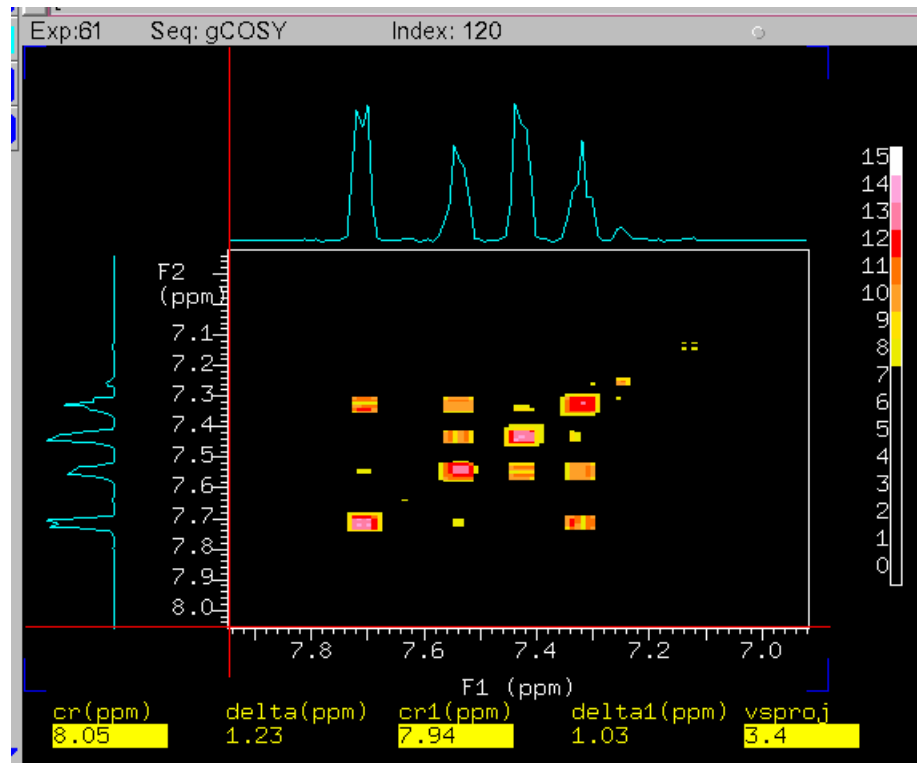


Figure 65. gCOSY Spectrum (Aromatic Region) from 10% 2-ethyl-1-indanone

### *Injection Method*

A Hamilton Gastight 25- $\mu$ L syringe was filled with clean  $\text{CDCl}_3$  solvent and used to rinse the probe. Then, 5- $\mu$ L of 10% 2-ethyl-1-indanone (519  $\mu$ g; 3.2  $\mu$ mol; v/v in  $\text{CDCl}_3$ ) was drawn into the syringe and injected into the probe. Using the same syringe, 12  $\mu$ L of clean  $\text{CDCl}_3$  was picked up and injected into the probe to push the 5- $\mu$ L sample into the NMR flow cell of the CapNMR probe. The 12- $\mu$ L Push Volume was calibrated in advance of sample injection.

### *Screenshots*

All of the screenshots shown in this Technical Bulletin are from a Varian Inova console running Varian VnmrJ 2.1B software on a Unix workstation. The magnet was an Oxford 600 MHz magnet. (For more information on the Varian VnmrJ software, see <http://www.varianinc.com/cgi-bin/nav?products/nmr/>)